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IN SITU CHEMICAL REACTIONS ON THIN LAYERS IN THE IDENTIFICATION OF ORGANIC COMPOUNDS

INTRODUCTION

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SUMMARY

The literature on the subject of in situ reactions on thin layers is reviewed and a simple scheme for classifying the techniques involved is put forward. The review deals with those organic reactions which aid identification of substrates, but not with reversible complexing reactions (e.g. with silver nitrate), where the applied material can be recovered unchanged. Spontaneous or unintended reactions are distinguished from organised reactions with which most of the review is concerned.

The idea of carrying out a reaction on chromatography paper and then separating the products of the reaction on the same piece of paper has long been known. It was therefore not a great step to using the same principle on thin-layer chromatoplates. The range of the technique has been considerably extended on thin layers, since the latter are much better suited to the more corrosive reagents often necessary.

In this review attention is confined to those cases involving organic substrates, where the reaction takes place in situ on the chromatoplate, though it is often possible to carry out the same reaction on a microgram scale in a small vessel and then to apply the reaction products to the chromatoplate. Reaction in situ, where it is possible to apply it, helps to preserve the advantage of speed possessed by TLC, though it may be somewhat lacking in degree of control compared with reaction in a flask or capillary. It should also be pointed out that, in this review, no attempt has been made to include reversible as well as irreversible reactions (e.g. complexing with silver nitrate, boric acid, etc., which are essentially an aid to separations), though they can play a significant role in the identification of substances.

Two main classes of reaction in situ on thin-layer plates may be distinguished, organised or intended reactions (O) and spontaneous or unintended reactions (S). Particular attention will be paid to organised reactions, since these are of particular value for the purpose of identification. However, it should be pointed out that good reproducibility in organised reactions can be jeopardised by the effects of simultaneous but unintended reactions.

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The first application of in situ reactions on thin layers to the identification of substances was probably that described by MILLER AND KIRCHNER¹ in 1953. Since then the technique in various modes has been applied to a wide variety of compounds, though the literature is not particularly extensive. A good deal of the work up to 1965 has been reviewed in a book by KIRCHNER², who covered reactions involving oxidation, reduction (or hydrogenation), dehydration, hydrolysis, esterification, bromination, enzyme action and the formation of certain derivatives. In the past few years the range of reactions has not been extended much, but there have been an appreciable number of new applications. Both one-dimensional (X1) and two-dimensional (X2) techniques have found new applications and the various types of techniques are summarised in Table I and in Fig. 1.

TABLE I CLASSIFICATION OF TECHNIQUES

Code	Method type		
O S D	Organised, intended Spontaneous, unintended Defined reaction	Reaction types	
F	Fingerprint method		
AR AG AL AM AS	Radiation, heat, light Gas, vapour Liquid spray In mobile phase In stationary phase	Methods of reactant application	
X ₁ X ₂ X ₂ RSS X ₂ SRS X ₂ RSRS M	One-dimensional Two-dimensional Two-dimensional with reaction before first development Two-dimensional with reaction between developments (diagonal chromatography) Two-dimensional with reaction before each development Miscellaneous	TLC	

One may broadly distinguish two types of reaction product. In the first case (D) the product is one of a few simple and identifiable compounds via a defined reaction as in, for instance, the formation of cinnamyl alcohol from cinnamaldehyde by reduction with lithium aluminium hydride¹. In the second case (F) the product is complex and unidentified as in the technique described by MATHIS AND OURISSON³ and by WILK AND BRILL⁴; this may be classed as a 'fingerprint' method and is the one less commonly used (see Fig. 1b).

A summary of the various reactions which have been reported as being possible to carry out *in situ* is given in Table II. In the paragraphs which follow the application of these reactions and techniques to particular problems is briefly discussed.

CARBOHYDRATES, PROTEINS AND AMINO ACIDS

There have been comparatively few reports on the application of reaction techniques to these classes of compound, PATAKI et al,^{5,0} have prepared the dinitro-

TABLE II
SUMMARY OF REACTIONS

No.	Reaction	Reference
I	Acetylation	3, 20, 21, 25, 27
2	Dehydration by H ₂ SO ₄	1, 26
3	Derivative formation	
	(a) Acetate	See reaction 1
	(b) Dinitrobenzoate	I
	(c) Dinitrophenyl (of amino acid)	5, 6
	(d) Dinitrophenylhydrazone	21, 25, 26, 29
	(e) Methyl ester	See reaction 5
	(f) Phenyl isocyanate	ī
	(g) Semicarbazone	I
4	Diazotisation	30, 31
5	Esterification of acids	20, 21
6	Halogenation	4, 10, 11, 18, 25, 28
7	Hydrogenation, catalytic	10, 11, 25
7 8	Hydrolysis with acid	14, 15, 16, 17, 33
9	Hydrolysis with alkali	1, 20, 25
10	Isomerisation	23
II	Methanolysis	12, 13, 16, 17
12	Nitration	25, 32
13	Oxidation with chromic acid	1, 24, 25, 26
14	Oxidation with nitric acid	24
15	Oxidation with oxygen, ozone	28
16	Oxidation with peroxide, peracid	3,8
17	Photochemical reaction	7, 22, 23
18	Pyrolysis	34
19	Reduction with FeSO ₄	18
20	Reduction with LiAlH ₄	r
21	Reduction with NaBH ₄	24,25
22	Reduction with Sn + HCl	31
	Reduction with hydrogen	See reaction 7
23	Miscellaneous	9, 25, 28

phenyl derivatives of amino acids in situ on silica gel by spraying with dinitrofluorobenzene reagent (method types O/D/AL/XI, X2 RSS and X2 SRS) and RIVETT AND WILSHIRE⁷ have studied the photochemical decomposition of certain short-chain peptides on silica gel (method types O/D/AR/XI, X2 RSS).

LIPIDS AND ASSOCIATED COMPOUNDS

Several reactions have been applied in the analysis and identification of fatty acids; Malins and Mangold⁸ have oxidised unsaturated methyl esters with peracetic acid (method type O/D/AL/XI); Johnson et al.⁹ have made use of the specific reaction with silver nitrate to identify and determine cyclopropene acids (method type O/D/AS/XI) and Kaufmann and coworkers^{10,11} have described the catalytic hydrogenation of unsaturated fatty acids on kieselguhr¹⁰ or gypsum¹¹ (method types O/D/AG/XI, X2 SRS). The same authors^{10,11} have also described the bromination of both fatty acids and triglycerides (method types O/D/AM/XI, X2 SRS).

The methanolysis by 12% KOH in methanol of phospholipids on silica gel has been described by KAUFMANN et al. 12 (method types O/D/AL/XI, X2 SRS) for both qualitative and quantitative analysis of mixtures and OETTE AND Doss 13 have

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compared quantitatively the 'in situ' and 'off plate' methanolysis of lipids (method type O/D/AL/XI).

Several papers have reported the use of *in situ* reaction for the identification and analysis of plasmalogens; Schmid and Mangold¹⁴ and Horrocks¹⁵ have described two-dimensional procedures for identification of plasmalogens and the aldehydes formed from them on acid hydrolysis (method type O/D/AG/X2 SRS). Recently a very interesting two-dimensional procedure, involving both hydrolysis of the plasmalogens and the methanolysis of the acyl phosphatide residues *in situ*, has been developed by Lundberg and co-workers^{16,17} (method type O/D/AG + AL/X2 RSRS).

SCOTNEY AND TRUTER¹⁸ have used *in situ* reduction of peroxides by ferrous salt (method type O/D/AL/X2 SRS) in order to identify the products of autoxidation of lanostenyl acetate. Copius Peereboom and Beekes¹⁹ have shown how cholesterol can be distinguished from its critical pair, brassicasterol, by development in solvent containing bromine (method type O/D/AM/XI).

The identification of aliphatic alcohols, esters and acids, using in situ acetylation, hydrolysis and esterification with methanol, has been described by Purdy AND Truter²⁰ (method type O/D/AL/XI); similar acetylation and esterification reactions have also been employed by Holloway and Challen²¹ in the identification of natural waxes (method type O/D/AL/XI).

SUBSTANCES OF PHARMACOLOGICAL INTEREST

A paper by STAHL²² concerning the effect of light and air on pyrethrins is the first describing the use of *in situ* reaction between developments in two dimensions, the method usually referred to as diagonal chromatography (also SRS or TRT technique) (see Fig. 1d). This method was later used by SCHUNACK AND ROCHELMEYER²³ to study the photochemical *cis-trans* isomerisation of annulolins on silica gel (method type O/D/AR/X2 SRS).

An important paper by Kaess and Mathis²⁴ discusses the reduction in situ by sodium borohydride and oxidations by nitric acid or by chromic acid in the identification of alkaloids (method type O/D/AL/XI); several other reactions in capillaries (before application to the chromatoplate) are mentioned in this same paper. The identification of alkaloids by reaction with iodine vapour, a 'fingerprint' method, has been described by Wilk and Brill⁴ (method type O/F/AG/XI).

A very useful paper by ELGAMAL AND FAYEZ²⁵ should also be mentioned; this describes methods for many *in situ* reactions on silica gel or alumina, as applied to a wide variety of model compounds of pharmacological interest (method type O/D/AL + AG/XI).

MISCELLANEOUS SUBSTANCES

The identification of certain terpene alcohols, employing oxidation by chromic acid, dehydration by sulphuric acid and the formation of DNPH derivatives of carbonyls, has also been described by Mathis²⁶ (method type O/D/AL/XI) and the author himself²⁷ has recently described the partial acetylation of polyglycerols in situ (method type O/D/AL/X2 SRS).

WILK et al.²⁸ have discussed the use of reaction with various gases or vapours (O₂, O₃, N₂O₃, NO₂, NO, Cl₂, Br₂, I₂, SO₂ and HNO₃) in a fingerprint method for the identification of various aromatic bases and polynuclear aromatic hydrocarbons (method type O/F/AG/XI) and the formation of DNPH derivatives of the bromophenacyl esters of some aromatic acids has recently been employed by Stedman²⁰ (method type O/D/AL/XI). The diazotisation of aromatic amines in situ on silica gel and the coupling of the diazo compounds with a phenol has been described by Marcus et al.³⁰ in a study of dye products from commercial amines (method types O/D/AL/XI, X2 SRS) and Thawley³¹ has shown that aromatic nitro compounds can be reduced on silica gel by tin and hydrochloric acid and the amine formed diazotised. Polesuk and Ma³² have also recently described the nitration of aromatic compounds on thin layers and a method for the partial hydrolysis of p-aminobenzoic esters and of sulphonamides on silica gel by hydrochloric acid vapour has been reported³³ (method type O/D/AG/XI).

Finally mention should be made of a new technique³⁴ involving TLC in one dimension of the pyrolysis products of a substance (not formed *in situ*); a pseudo two-dimensional modification, with the exit port of the pyrolysis chamber being moved along one axis of the chromatoplate as the pyrolysis temperature increases, is likely to give a two-dimensional pattern, which is highly characteristic of any given substance, provided it is reproducible.

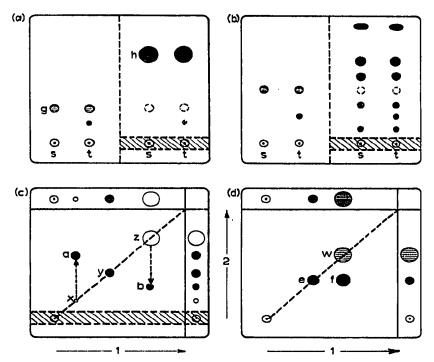


Fig. 1. (a) One-dimensional organised defined reaction (type O/D/X1). s= standard, t= test material. h is formed from g (e.g. a carbonyl by oxidation of an alcohol). (b) One-dimensional organised fingerprint reaction (type O/F/X1). s= standard, t= test material. (c) Two-dimensional organised defined reaction (type O/D/X2 SRS). Reagent is applied to shaded area between developments; a is formed from x, y does not react and b is formed from z. (d) Two-dimensional spontaneous defined reaction (type S/D/X2 RSRS); e is formed from w before first development and f is formed similarly between developments in the same solvent, if e=f. Hatched spot, original substance; black spot, product from in situ reaction; the shaded area is the area to which reagent is applied.

SPONTANEOUS REACTIONS

It is not the author's intention to review also the subject of the spontaneous or unintended reaction in situ (type S), but some mention of it seems necessary. Many of the secondary reactions that can be caused by the adsorbent have been reviewed by Lederer and Lederer³⁵ and there have been a number of recent publications dealing with instability of lipids³⁶⁻³⁸, of olefines³⁹, of steroids⁴⁰⁻⁴², of carotenoids⁴³, of pyrazolones⁴⁴, aflatoxins⁴⁵, and silyl ethers⁴⁶ on thin layers, due to the action of the adsorbent, of light or of atmospheric oxygen.

As Penner et al.⁴² point out, the appearance of several spots on a normal chromatoplate and the conclusion that the original substance is impure may be an erroneous one. It must therefore be necessary to take care to ensure that the products of an organised reaction in situ are the result only of that reaction on the substrate and not partly the result of a spontaneous reaction. Stahl⁴⁷ and others⁴² have clearly described how one may detect instability of a compound on a chromatoplate using a simple two-dimensional procedure (see Fig. 1d).

REFERENCES

- 1 J. M. MILLER AND J. G. KIRCHNER, Anal. Chem., 25 (1953) 1107. 2 J. G. KIRCHNER, Technique of Organic Chemistry, Vol. 12, Thin-layer Chromatography, Interscience, New York, 1967. 3 G. MATHIS AND G. OURISSON, J. Chromatog., 12 (1963) 94. 4 M. WILK AND U. BRILL, Arch. Pharm., 301 (1968) 282. 5 G. PATAKI, J. Chromatog., 16 (1964) 541.
 6 G. PATAKI, J. BORKO, H. CH. CURTIUS AND F. TANCREDI, Chromatographia, 1 (1968) 406. 7 D. E. RIVETT AND J. F. K. WILSHIRE, Photochem. Photobiol., 6 (1967) 867.

 8 D. C. Malins and H. K. Mangold, J. Am. Oil Chemists' Soc., 37 (1960) 576.

 9 A. R. Johnson, K. E. Murray, A. C. Fogerty, B. H. Kennett, J. A. Pearson and F. S. Shenstone, Lipids, 2 (1967) 316.

 10 H. P. Kaufmann, Z. Makus and T. H. Khoe, Fette Seifen Anstrichmittel, 64 (1962) 1. 11 H. P. KAUFMANN AND T. H. KHOE, Fette Seifen Anstrichmittel, 64 (1962) 81. 12 H. P. KAUFMANN, S. S. RADWAN AND A. K. S. AHMAD, Fette Seifen Anstrichmittel, 68 (1966) 13 K. OETTE AND M. Doss, J. Chromatog., 32 (1968) 439. 14 H. H. O. SCHMID AND H. K. MANGOLD, Biochim. Biophys. Acta, 125 (1966) 182. 15 L. A. HORROCKS, J. Lipid Res., 9 (1968) 469.
 16 C. V. VISWANATHAN, M. BASILIO, S. P. HOEVET AND W. O. LUNDBERG, J. Chromatog., 34 (1968) 241.

 17 C. V. VISWANATHAN, M. BASILIO, S. F. HUEVET AND W. C. LONDBERG, J. Chromatog., 38 (1968) 267.

 18 J. SCOTNEY AND E. V. TRUTER, J. Chem. Soc., (1968) 1911.

 19 J. W. Copius Peereboom and H. W. Beekes, J. Chromatog., 9 (1962) 316.

 20 S. J. PURDY AND E. V. TRUTER, Proc. Roy. Soc. (London), Ser. B, 158 (1963) 536 and 544.

 21 P. J. HOLLOWAY AND S. B. CHALLEN, J. Chromatog., 25 (1966) 336. 22 E. STAHL, Arch. Pharm., 293 (1960) 531. 23 W. SCHUNACK AND H. ROCHELMEYER, Arch. Pharm., 298 (1965) 572. 24 A. KAESS AND C. MATHIS, Ann. Pharm. Franc., 24 (1966) 753. 25 M. H. A. ELGAMAL AND M. B. E. FAYEZ, Z. Anal. Chem., 226 (1967) 408. 26 C. Mathis, Ann. Pharm. Franç., 23 (1965) 331. 27 M. S. J. DALLAS, J. Chromatog., 48 (1970) 225. 28 M. WILK, U. HOPPE, W. TAUPP AND J. ROCHLITZ, J. Chromatog., 27 (1967) 311. 29 E. D. STEDMAN, Analyst, 94 (1969) 594.
 30 B. J. MARCUS, A. FONO AND T. S. MA, Mikrochim. Acta, (1967) 960.
 31 A. R. THAWLEY, J. Chromatog., 38 (1968) 399.
 32 J. POLESUK AND T. S. MA, Mikrochim. Acta, (1969) 352.
- J. Chromatog., 48 (1970) 193-199

33 M. BAGGIOLINI AND B. DEWALD, J. Chromatog., 30 (1967) 256. 34 R. N. ROGERS, Anal. Chem., 39 (1967) 730.

35 E. LEDERER AND M. LEDERER, Chromatography, Elsevier, Amsterdam, 1957, Ch. 8.

- 36 J. J. Wren and A. D. Szczepanowska, J. Chromatog., 14 (1964) 405. 37 I. Mitsev, J. Slavceva and A. Popov, C.R. Acad. Bulg. Sci., 20 (1967) 693.
- 38 K. IMAICHI, J. FUKUDA AND A. NOTOMI, J. Biochem., 59 (1966) 189.
- 39 S. LANDA AND L. MARKOVEC, Collection Czech. Chem. Commun., 29 (1964) 2309.
- 40 D. R. IDLER, N. R. KIMBALL AND B. TRUSCOTT, Steroids, 8 (1966) 865.
 41 M. TALAAT ABDEL AZIZ AND K. I. H. WILLIAMS, Steroids, 12 (1968) 167.
- 42 M. H. PENNER, J. M. TALMAGE AND M. GELLER, J. Pharm. Sci., 55 (1966) 429.
- 43 J. W. NELSON AND A. L. LIVINGSTON, J. Chromatog., 28 (1967) 465.
- 44 K. MACEK, J. Chromatog., 33 (1968) 332.
- 45 T. W. KWON AND J. C. AYRES, J. Chromatog., 31 (1967) 420.
- 46 J. LEHRFELD, J. Chromatog., 32 (1968) 685.
- 47 E. STAHL, Thin-layer Chromatography, Springer Verlag, Berlin and Academic Press, New York.

DISCUSSION

DE ZEEUW: When diagonal chromatography (separation-reaction-separation procedures, S-R-S) is applied, the nonreacting substances are expected to lie on the diagonal if the same solvent is used for development. However, is there an influence due to the direction in which the layer was spread and is it possible that when working with complex mixtures displacement will occur in the direction of the first development?

DALLAS: The position of the spots can vary very slightly with the direction of development, but I have found this unimportant in practical interpretation of the two-dimensional chromatograms. The effect of overloading on the distortion from the true diagonal is more important from my experience.

HAIS: If the position of the spot is modified in the first dimension by the presence of other components of the sample, especially if displacement occurs, then in the second dimension the substance may move without being affected (displaced) by other components, from which it was already separated in the first run. This will result in off-diagonal spots and should be borne in mind in the interpretation of the pattern.

1. Chromatog., 48 (1970) 193-199